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# 13. Introduction to the Scaling of Aquatic Animal Locomotion

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## ABSTRACT

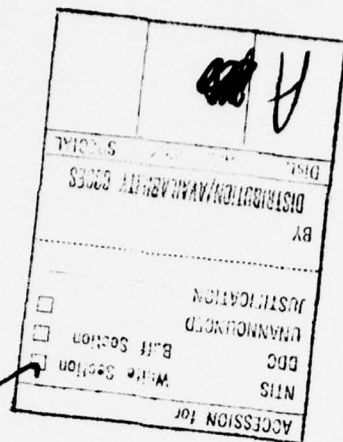
This introductory lecture attempts to discuss the scaling problems related to the locomotion of aquatic animals in two major categories characterized by low and high Reynolds numbers of swimming motion. In the low-Reynolds-number flow regime it first gives a brief survey of flagellar and ciliary locomotion in order to ascertain the different key parameters that play a role. The hydromechanical and physiological performance of a group of micro-organisms, of different sizes but otherwise similar in their organization from the scaling point of view, is examined.

In the high-Reynolds-number category, discussions of the scaling problems include the carangiform and lunate-tail locomotion of different groups of fishes and cetaceans, with consideration of hydromechanical efficiency and physiological function. The discussion is based on the data of both comparative zoology and dynamical similarity.

## INTRODUCTION

It is most gratifying to see this Cambridge Symposium take on the main theme of *scaling and similitude* in animal locomotion. Its importance for the future development of biological science has become increasingly recognized. A timely exploration of this theme, which is the objective of this meeting, will certainly add strength to and point out new, rewarding directions for current research activities such as those presented at the International Symposium on Swimming and Flying in Nature held in Pasadena in July 1974.

It is a great privilege to have been called upon by Professor Lighthill and Professor Weis-Fogh to give an introductory talk on the influence of size on the different types of aquatic locomotion found in various groups of animals.



In accepting the call I fully realize that the total range of subject-matter to be covered is very extensive and it would be hard to emulate Professor Lighthill's elucidation of the scaling aspects of some 10 groups of aquatic animals at the 1973 Duke University Conference. I feel, nevertheless, that I have benefited from this exciting challenge, for already I have found that important and difficult problems in fluid mechanics have been illuminated by studies in metabolism and chemical energy conversion, and vice versa. For example, we can actually make a better estimate of the drag of a fish from biological measurements than from direct flow measurement, because the latter is formidably hard. This underlines the importance of close collaboration between scientists from different fields engaged in interdisciplinary research on topics such as animal locomotion. In this regard I would like to acknowledge with appreciation the co-operation of the two groups at Cambridge, under Sir James Lighthill and Professor Torkel Weis-Fogh respectively, as well as that of many other colleagues from all over the world.

I would like to discuss the scaling problems of aquatic animal locomotion in two major categories, characterized respectively by low and high Reynolds numbers of the swimming motion, which are broadly equivalent to aquatic animals of microscopic and macroscopic size. In the low-Reynolds-number regime I shall give a brief survey of flagellar and ciliary propulsion in order to ascertain the key parameters underlying the hydromechanical and physiological performance of the two groups of micro-organisms. Here the limited range of animal sizes, swimming speeds, and beat frequencies observed in specific groups, coupled with the very limited data available on their energy conversion processes, show that a lot more information is required before a good understanding can be reached.

In the high-Reynolds-number category, I shall concentrate on the scaling problems of carangiform and lunate-tail locomotion of different groups of fishes and cetaceans, considering their hydromechanical efficiency and physiological functions on the basis of data from both comparative zoology and dynamical similarity. This is an area where experimental results are relatively abundant, and there has recently been considerable research activity, so that a more refined investigation of the scaling problem is possible. Current studies combining both physiology and dynamics promise further progress which may have far-reaching effects, as I hope to point out later.

#### LOCOMOTION OF MICRO-ORGANISMS

The two major kinds of microscopic organisms to be discussed here are the flagellates and the ciliates. In general they have a maximum linear dimension smaller than 1 mm, with a majority of them being much smaller. Flagellates

propel themselves through an aqueous medium by performing undulatory movements of a long thin flagellum (or several flagella) in the form of a planar, helical, or a general three-dimensional wave. Ciliary propulsion utilizes the co-ordinated effort of a large number of flagella-like organelles, called cilia, which are distributed over the cell membrane, and wave in a manner known as 'metachronism'. Both flagellates and ciliates swim in aqueous media, attaining a characteristic velocity,  $V$ , in such a way that the Reynolds number

$$Re = Vl/\nu \quad (1)$$

is very low. Here  $\nu = 0.01 \text{ cm}^2 \text{ s}^{-1}$  is the kinematic viscosity of water, and  $l$  is the length of the organism. Typical values of the Reynolds number are around  $10^{-3}$  for spermatozoa,  $10^{-6}$  for most bacteria, and in the range  $0.01 < Re < 1$  for various ciliates (the Reynolds number based on ciliary length  $l_c$  is considerably smaller). At these low Reynolds numbers, all the inertial effects, both of the liquid and of the organism, become insignificant. Consequently, hydromechanical calculations of flagellar and ciliary propulsion are generally based on neglecting the inertial effects, leaving the viscous stress and pressure as the only quantities that need be considered in the momentum balance for given swimming movements.

From the laws of motion, the net momentum and net angular momentum of the fluid surrounding a self-propelling micro-organism, swimming at constant velocity, must be zero. However, the backward drag force and the equal and opposite thrust acting on a swimming micro-organism cannot be separated, as they can for a large aquatic animal (as we shall see), and must be dealt with together. For a long, thin organelle such as a flagellum or a cilium, the forces tangential and perpendicular to its centerline, denoted by  $F_s$  and  $F_n$  respectively, which it exerts on the fluid while moving with tangential and normal velocities  $V_s$  and  $V_n$ , depend mainly on its length  $l$  and not on its transverse dimensions. Calculations of these forces can be considerably simplified by means of the "resistive-force theory", which was first developed for flagellar applications by Hancock<sup>(33)</sup> and Gray and Hancock,<sup>(31)</sup> was extended to helical movements by Chwang and Wu,<sup>(25)</sup> and was recently refined further by Lighthill.<sup>(48,49)</sup> It provides a simple relationship between the force and the velocity in the form

$$F_s = C_s V_s l, \quad F_n = C_n V_n l, \quad (2)$$

where  $C_s$  and  $C_n$  are the resistive force coefficients which depend on the transverse dimensions and wavelength of the body but not on its velocity. Both  $C_s$  and  $C_n$  are proportional to the (dynamic) viscosity coefficient of the fluid,  $\mu$  (equal to the kinematic viscosity,  $\nu$ , times the density  $1 \text{ g cm}^{-3}$ ).



The ratio of  $C_s$  to  $C_n$  or

$$\gamma = C_s/C_n \quad (3)$$

plays an important role in swimming propulsion since its value (always  $< 1$ ) determines what type of undulatory motion will give the optimum forward velocity or optimum efficiency, as first elucidated by Lighthill (Refs 43, p. 442; 48, p. 122). I shall try to recapitulate his argument by considering the ideal case of *zero-thrust swimming* of a flagellum (such as that of a sperm detached from the cell body, or of a spirochete). I shall deal solely with the translational motion, leaving the possible angular motion and all further details to the literature. (The same argument can be applied, qualitatively at least, to larger elongated animals.)

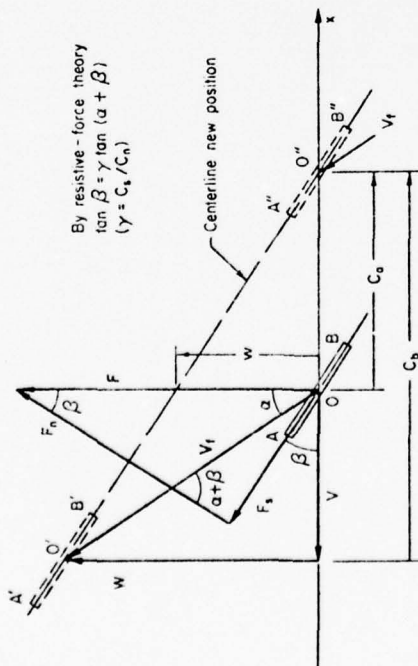


FIG. 1. A resistive-force theory diagram. A segment AB of an elongated body in undulatory motion moves with velocity  $V_f$  and exerts a force  $F$  on the fluid in a zero-thrust propulsion. Its lateral velocity is  $W$  relative to the animal's mean position and  $w$  relative to the water. The wave velocity is  $c_s$  relative to the water and  $c_b$  relative to the body.

Figure 1 shows a segment AB of an elongated animal supposed to be performing a plane or helical wave, and achieving a swimming velocity  $V$  in the negative x-direction. In the ideal case of zero thrust (or zero x-component force), the force  $F$  exerted by the segment on the fluid can only be in a transverse direction as shown. Nevertheless there is a forward tangential component  $F_s$  along the centerline of the body when the latter is inclined to the plane of swimming at an angle  $\beta$ . (Figure 1 may be envisaged as in the plane of a planar wave, or, in the case of a helical wave, in the plane tangential to the helical movement of the segment in question.) The segment, under the

combined traction of  $F_s$  and the normal component of force,  $F_n$ , will move to a new position  $A'B'$  with velocity  $V_f$  which has the forward x-component  $V$  and a transverse component  $W$ . Clearly, we have

$$F_s = F \sin \beta, \quad F_n = F \cos \beta, \quad (4)$$

and

$$V_s = V_f \sin(\alpha + \beta), \quad V_n = V_f \cos(\alpha + \beta), \quad (5)$$

where  $\alpha$  is the angle between  $F$  and  $V_f$ . Substituting (4) and (5) in (2) yields

$$\tan \beta = \gamma \tan(\alpha + \beta). \quad (6)$$

The angle  $\alpha$  vanishes when  $\beta = 0$  or  $\pi/2$ , and achieves a maximum when

$$\beta = \beta_m = \arctan \gamma^{1/2}, \quad (7)$$

giving

$$V_{\max}/W = \tan \alpha_{\max} = \frac{1}{2}(1 - \gamma)^{1/2}. \quad (8)$$

In the meantime the constant phase of the body wave has been propagated backwards to reach position  $A''B''$  (which would now be occupied by a new segment of the body) with wave (phase) velocity  $c_s$  relative to the water, or with wave velocity

$$c_b = c_s + V \quad (9)$$

in the body frame of reference. Since  $W/c_b = \tan \beta$  (see Fig. 1),

$$V_{\max}/c_b = \tan \alpha_m \tan \beta_m = \frac{1}{2}(1 - \gamma). \quad (10)$$

The lateral velocity of the segment relative to the water,  $w$ , is equal to  $W(c_s/c_b)$ , which when  $\beta = \beta_m$  assumes a value such that

$$\frac{w}{W} = \frac{c_s}{c_b} = \frac{1}{2}(1 + \gamma). \quad (11)$$

The above results agree with Lighthill's (Ref. 48, p. 123). An important consequence is that for fixed  $W$  (or  $c_b$ ),  $V_{\max}$  decreases with increasing  $\gamma$ , and vanishes at  $\gamma = 1$ , which means self-propulsion in the present case (of zero thrust) would be impossible when  $C_s = C_n$ .

A more general situation arises when a flagellum or a ciliary system has a passive cell body to propel so that each segment must on average have its force  $F$  directed both sideways and somewhat forward. The efficiency may then be defined as the ratio of useful work done by the forward component of  $F$  in propelling the animal forward at speed  $V$ , to the power required which is  $F \cdot V_f$ . The values of  $V_{\max}$  and  $V$  at the maximum efficiency can also be

calculated. Lighthill<sup>(48)</sup> showed that the value of  $V$  at maximum efficiency ( $\eta = \eta_{\max}$ ) is

$$V/c_b = 1 - \gamma^{1/2} \quad \text{and} \quad \eta_{\max} = (1 - \gamma^{1/2})^2. \quad (12)$$

This value of  $V$  is not much less than  $V_{\max}$  (Eqn 10), even when  $\gamma = 0.2$ .

Detailed analysis has produced the following expressions for the force coefficients in the low Reynolds number case

$$C_s = \frac{2\pi\mu}{\ln(2q/b) - 1/2}, \quad C_n = \frac{4\pi\mu}{\ln(2q/b) + 1/2}, \quad (13)$$

where  $b$  is the cross-sectional radius of the thin segment, and  $q$  is a quantity depending on body shape and certain wave parameters. Lighthill<sup>(49)</sup> contended that for flagellar waves with wavelength  $\lambda$  it is accurate to take

$$q = 0.09\lambda, \quad (14)$$

and to delete the term  $-1/2$  in the denominator of the expression for  $C_s$ . (The values of  $q$  for rigid slender bodies of various shapes are given by Batchelor,<sup>(8)</sup> Tillet,<sup>(61)</sup> Cox<sup>(26)</sup>). The value of  $\gamma$ , according to these formulas, lies in a range between 0.6 and 0.7 and is not close to 0.5 as earlier theories have commonly assumed. The corresponding efficiency is therefore about 0.05 for flagellar propulsion, which is rather low, and  $V/c_b \approx 0.2$ . Lighthill<sup>(46)</sup> noted that for swimming snakes  $\gamma$  is likely to be about 0.1, leading to  $\eta \approx 0.5$  and  $V/c_b \approx 0.7$ .

Another important feature of flagellar propulsion is that the maximum inclination of a plane flagellar wave, or the pitch angle of a helical flagellar wave, should, for maximum speed, be

$$\beta_{\max} \approx 40^\circ \quad (15)$$

(from Eqn 7) for  $\gamma$  around 0.6. This figure is close to the value for minimum power output, and comparable values are indeed observed.

With this introduction we now proceed to discuss the problems of flagellar and ciliary propulsion separately.

### Flagellar Propulsion

The mechanical power needed for a flagellar organism of length  $l_f$  to swim at velocity  $V$  may be readily estimated by applying the resistive theory, giving

$$P_f = F_s V_s + F_n V_n = C_n (\gamma V_s^2 + V_n^2) l_f \approx 50\mu V^2 l_f, \quad (16)$$

where the constant factor 50 is borne out by detailed calculation to be typical of flagellar movement. It is based on the observation that there is not much difference in diameter (about  $0.2 \mu\text{m}$ ) between the flagella of different

species (excluding the bacterial flagella), and  $\lambda/b$  is typically between 100 and 200, though the length  $l_f$  may vary considerably from one species to another. Additional power is further required to overcome the drag of a passive cell body (of a three-dimensional form with length  $l_b$  say), if any, which is

$$P_b = \text{const.} \cdot \mu V^2 l_b. \quad (17)$$

The value of  $P_b$  is generally smaller than that of  $P_f$ .

To proceed with a scaling analysis we need to know the chemical energy input per flagellar beat (say  $E_f$ ); this can in principle be obtained from the measured rate of dephosphorylation of ATP (the chemical compound adenosine triphosphate) by the flagellum. Such information, however, is extremely scarce. Under the circumstances it has been suggested (see Alexander,<sup>(1)</sup> p. 40) that for protozoic flagella having the well-known arrangement of 9 + 2 pairs of micro-fibrils running along them, the maximum chemical energy input of a flagellum is likely to be proportional to its length  $l_f$  (or,

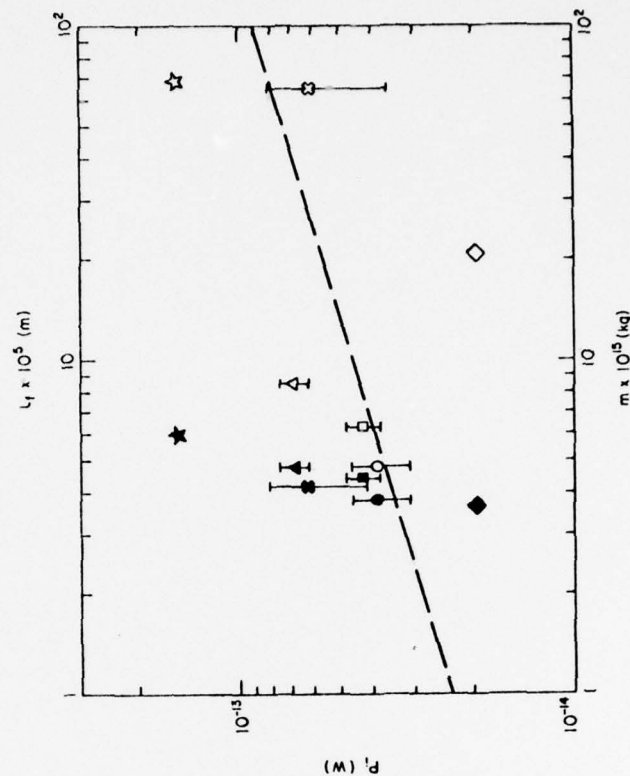


FIG. 2. Variation of chemical power input  $P_f$  of flagellates with their flagellar length  $l_f$  (in solid symbols) and mass  $m$  (in open symbols).  $\times$ —*Colobocentrotus atrajus*;  $\square$ —*Strongylocentrotus purpuratus*;  $\triangle$ —*Lytechinus pictus*;  $\diamond$ —*Ciona intestinalis*;  $\star$ —*Chaetopterus variopodatus*;  $\star$ —Bull sperm. Experimental data from Brokaw and Gibbons.<sup>(124)</sup> The slope of the dotted regression line is 0.28.

equivalently, to the mass of the flagellum since its cross-sectional diameter is uniform and constant). From this assumption and Eqn (16) it would follow that the swimming velocity  $V$  is scale-independent.

The experimental data which have made this preliminary study possible are taken from Brokaw and Gibbons<sup>(24)</sup> who investigated the process of converting the chemical energy of ATP into the mechanical work of sliding between the doublet tubules of the axoneme, and hence of generating particular flagellar waves, in six different species of spermatozoa. Figure 2 shows the dependence of chemical power input of the flagellates,  $P_f$ , on the flagellar length  $l_f$  (which is 3 to 15 times the cell-body length) and the total mass  $m$  of the organism. Although a regression line can be drawn by the method of least squares for the  $\log P_f - \log m$  relation, with slope 0.28, the data are considered to be too scattered for a confident conclusion to be drawn. The short range of flagellar length  $l_f$  covered by these species makes determination of the  $P_f - l_f$  relation difficult, but, from these results at least,  $P_f$  is clearly far from being proportional to  $l_f$  as predicted.

The difficulties seem to stem from the apparent lack of correlation between  $l_f$  and  $m$ , as can be seen from Fig. 2. This is in fact not surprising since the thrust required from each segment of a long flagellum to balance the drag force on the cell body is very small. Thus changes in mass, and hence drag force, will involve only very slight modification of the propulsive mechanism, which will still be approximately described by the zero-thrust theory outlined above. This implies that the flagellar length  $l_f$  (or the total cell length  $l$ ) may not be the pertinent parameter to use in scaling; perhaps future experimental results will help settle this point.

We present in Fig. 3 the variation of swimming velocity  $V$  with  $m$ , for which a least square fit yields  $V = \text{const. } m^{-0.04}$ , i.e.  $V$  is virtually independent of body mass. This is in sharp contrast with the case of large animal locomotion, as we shall see in the discussion of the scaling of fish swimming.

The nondimensional quantity

$$\mathcal{E} = P/mgV, \quad (18)$$

where  $P$  stands for either the chemical power input or the mechanical power required, and  $g$  is the acceleration of gravity, provides a useful measure of the relative merit of different propulsive systems. If an animal expends power  $P = dE/dt$  to keep moving along a straight level path at speed  $V = dx/dt$ , where  $x$  is distance measured along the path, the ratio  $P/mgV = (mg)^{-1} dE/dx$  therefore gives the energy expenditure to transport unit weight unit distance. The parameter  $\mathcal{E}$  may thus be called the "specific energy cost" of transport (cf. the "net cost of transport" as defined by Alexander). It has been used by von Kármán and Gabrielli<sup>(66)</sup> to evaluate the comparative merits of 14 classes of transportation vehicle, and by Schmidt-Nielsen,<sup>(57)</sup> Tucker,<sup>(63, 65)</sup>

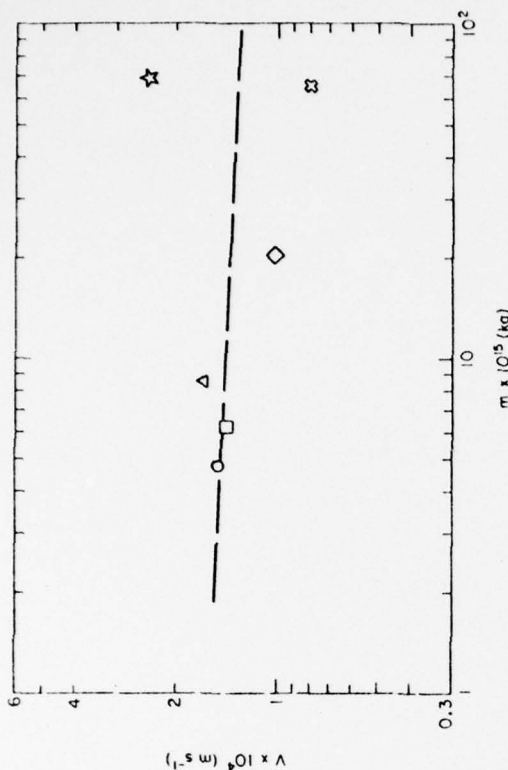


FIG. 3. Variation of swimming velocity of flagellates with their mass. The slope of the dotted regression line is  $-0.04$ . For other legends see caption to Fig. 2.

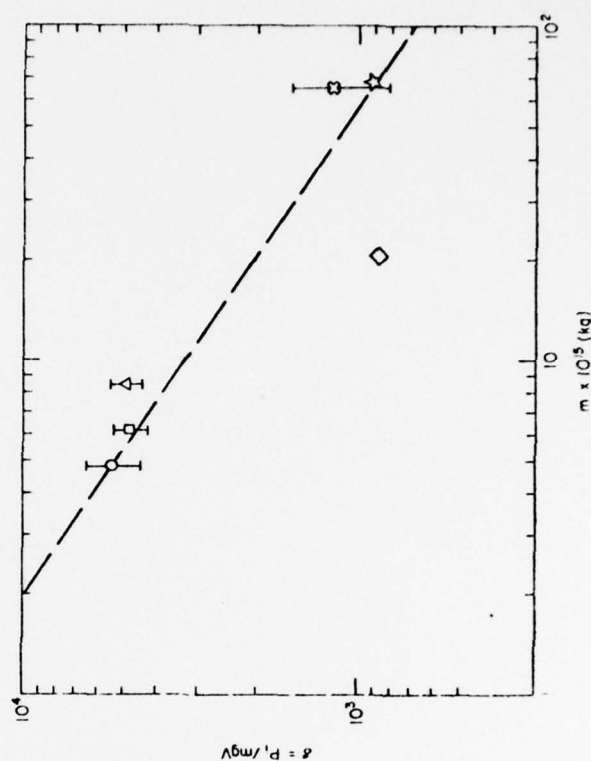


FIG. 4. The specific energy cost of flagellates versus their mass. The slope of the dotted regression lines is  $-0.68$ . For other legends see caption to Fig. 2.



and others for studies of comparative physiology. Figure 4 shows the specific energy cost of the six flagellates in question, which may be represented, by a least-square fit, as  $\mathcal{E} = \text{const. } m^{-0.68}$ . This  $\mathcal{E}$ - $m$  relationship is quite similar to those of other aerial, aquatic and terrestrial animal groups, but the slope  $-0.68$  of the regression line is steeper than most of them already known (see Tucker<sup>(65)</sup>).

Summarizing this preliminary study of flagellar propulsion, we have found

$$P_i \approx \text{const. } m^{0.28}, \quad V = \text{const. } m^{-0.04} \quad (19a)$$

and

$$\mathcal{E} = P_i/mg = \text{const. } m^{-0.68}. \quad (19b)$$

In view of the difficulties involved in such metabolism experiments, we should appreciate the value of these experimental results, although only six species have been studied, and not too much confidence can be placed in the quoted slopes of the regression lines. The results do throw enough light to make the following theoretical argument attractive.

Suppose we accept the assumptions that (i) the propulsive velocity  $V$  of a flagellum under the zero-thrust condition depends only on the wave form and beat frequency and is not appreciably affected by the presence of a cell body, and (ii) the power  $P_f$  required for maintaining a steady flagellar motion (hence  $P_f = \text{const.}$ ) forms a basal reference above which the power  $P_b$  required for propelling the cell body (Eqn 17) will vary when the cell body size  $l_b$  is varied. Then the changes in the power input,  $P_i$ , would scale according to

$$P_i \propto l_b \propto m^{1/3}, \quad \text{and hence} \quad \mathcal{E} = P_i/mgV \propto m^{-2/3}. \quad (20)$$

These relationships are in close agreement with (19a) and (19b). It will be of interest to acquire more experimental data for further studies along this line.

#### Ciliary Propulsion

Blake and Sleight<sup>(13)</sup> and Holwill<sup>(34a, b)</sup> have given excellent surveys upon the mechanics and physiology of ciliary propulsion, and their introductory lectures on the subject at the 1974 Caltech Symposium were of especial value. They pointed out that ciliary length, pattern of movement and rate of beat are important factors in determining the effectiveness of propulsion of fluids by cilia, whether for ciliated protozoa or metazoa, or for the inner organs of higher animals. Ciliates are generally known as very impressive swimmers when judged by their specific speeds (body lengths travelled/s). However, the four types of metachronal pattern in which ciliary movements are organised

(symplectic, antiplectic, dextiolectic, and laeoplectic) are so different from each other that it would be inappropriate to study ciliary efficiency collectively, as these authors and others have pointed out. This is also true for studies of scaling problems in ciliary propulsion.

Theoretical analysis of ciliary propulsion began when Taylor<sup>(60)</sup> introduced the "envelope model", based on the idea that the concerted action of large numbers of cilia on the fluid can be represented by the waving motion of an impermeable inextensible sheet enveloping the cilia tips. This model has been extended to cover an extensible sheet, with an estimate of the inertia effect by Tuck,<sup>(62)</sup> and has been applied by Blake<sup>(10, 11)</sup> to evaluate the motion of *Opalina*. It was pointed out by Lighthill<sup>(46)</sup> that it is only to symplectic motion that such a model should really be applied. In order to treat other types of metachronism, especially those such as antiplectic motion, in which the cilia tips may be far apart during the motion, Blake<sup>(12)</sup> developed a sublayer model in which each cilium was modelled by an elongated body attached to a flat cell surface.

Another simplified model, called the "traction-layer" model, has been investigated, at the suggestion of Wu,<sup>(77)</sup> by Keller.<sup>(39)</sup> It is based on the concept that the discrete forces of a system of cilia can be represented by an "equivalent" continuous distribution of an unsteady body force within the volume of the ciliary layer, provided that the spacings between adjacent cilia are small compared with the ciliary length,  $l_c$ . Conversion from the discrete to the continuum force is based on the resistive theory which gives the relationship between the original force exerted by a cilium, and its velocity relative to the flow produced by the ciliary system as a whole.

Using this procedure the model has been applied by Keller to an infinite plane ciliary layer with the body force represented by a Fourier series

$$f(x, y, t) = \sum_{n=0}^N f_n(y) \exp[in(kx - \omega t)] \quad (-\infty < x < \infty, 0 < y < l_c) \quad (21)$$

where  $N \leq \infty$ ,  $\lambda = 2\pi/k$  is the metachronal wavelength,  $\omega$  is the radian frequency, and  $f$  vanishes at the cell surface ( $y = 0$ ) as well as outside the cilia layer ( $y \geq l_c$ ). The functions  $f_n(y)$  can be given a polynomial representation or be determined by numerical methods. The Stokes equation with this force term included can be solved to give the flow velocity for particular movements of the cilia. A related optimisation problem is to find the external mean force distribution  $f_0(y)$  that will give the minimum power required,  $P_0$ , to maintain a rate of working on the fluid which is less than  $P_0$  by a fixed amount; this is equivalent to finding the minimum power required for fixed mean square of  $f_0$ , averaged across the cilia layer. The result for a four-term polynomial representation of  $f_0(y)$  is shown in Fig. 5. Thus we find the



mean flow velocity at the outer boundary of the cilia layer as

$$u_0(l_c) = 0.52A(\omega l_c)(kl_c) = 0.52Ac(2\pi l_c/\lambda)^2, \quad (22)$$

where  $c = \omega/k$  is the metachronal wave velocity and  $A$  is proportional to the amplitude of the dimensionless force distribution. The slope of  $u_0(y)$  at  $y = 0$  shows that there is a viscous skin friction at the cell surface which is counterbalanced by the fluid force acting on the cilia over the same area of the surface.

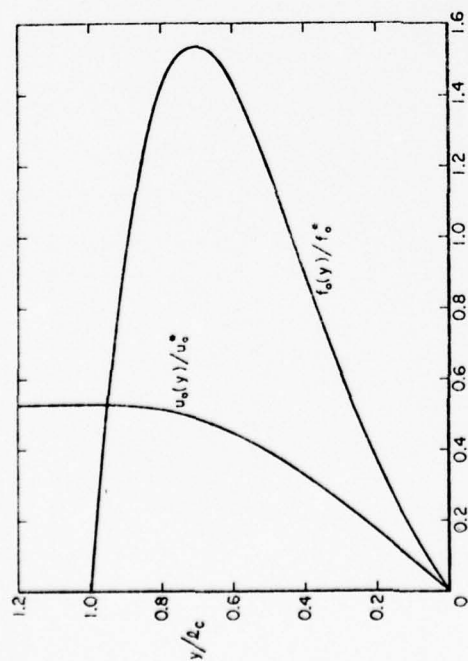


FIG. 5. The optimum continuous force distribution giving minimum power required for fixed mean square force, and the corresponding velocity profile.  $f_0^* = A\omega\omega/k^2$  and  $u_0^* = A\omega k l_c^2$  where  $l_c$  is the ciliary length,  $\omega$  is the radian frequency and  $k$  the wave number of the ciliary wave, and  $A$  is a constant making the  $y$ -integral of  $(f_0/f_0^*)^2$  unity.

The first harmonic velocity profile,  $u_1(y)$ , has been evaluated by Keller<sup>(39)</sup> in a few cases for which the wave parameters are available. The results for *Paramecium multimicronucleatum* are given in Fig. 6, from which we note that the contribution from the second harmonic is already very small. We further note from Fig. 6 that like the mean velocity  $u_0(y)$ ,  $u_1(y)$  is also proportional to  $(kl_c)^2$  or  $(l_c/\lambda)^2$ .

Experimental investigation of the scaling relation (22) has not yet been fully carried out. Furthermore, a complete examination of the scaling of ciliary propulsion cannot be made without physiological data on the chemical energy input, which are also not yet available. In addition, the hydromechanical analysis must be carried out for a finite shape, in the manner indicated by Blake<sup>(10)</sup> and Brennen.<sup>(17)</sup>

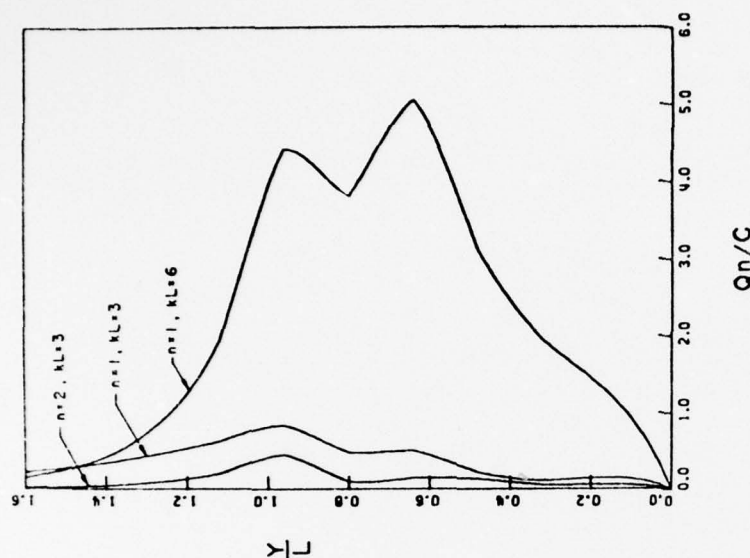


FIG. 6. Amplitude profile for the first and second harmonic  $x$ -components of flow velocity ( $Q_n$ , non-dimensionalised with respect to the metachronal wave speed  $c = \omega/k$ ) for *Paramecium* with  $ka = 0.80$  and  $kb = 1.60$ . ( $a$  and  $b$  are the row and column spacings between cilia, and  $L$  here denotes ciliary length).

## LOCOMOTION OF LARGE AQUATIC ANIMALS

The swimming motions of large aquatic animals are characterized from the hydrodynamicist's point of view by large values of the ratio of typical inertial forces to typical viscous forces in the surrounding water, i.e. of the Reynolds number,  $Re = V/\nu \gg 1$ . Here  $V$  is the mean swimming speed,  $l$  the length of the animal, and  $\nu$  the coefficient of kinematic viscosity of the fluid. In general, the Reynolds number lies in the range  $10^4 < Re < 10^8$  for various fishes and cetaceans swimming normally. At such high Reynolds numbers the effects of viscosity are mostly confined to a thin boundary layer adjacent to the body surface. This is especially true for the streamlined body shapes of

such animals in undulatory swimming motion, when flow separation is not observed to occur, and only a very thin wake is formed behind the body. In such cases the boundary layer will continually grow with distance along the fish, but its maximum thickness (at the tail end) is generally not more than a few per cent of the body thickness. Viscous effects can therefore be neglected in analysing the flow outside this boundary layer. The pressures acting on the body are those predicted by inviscid theory, because the boundary layer is too thin to support a pressure gradient across it. These pressures generate a thrust force which must overcome the drag force in order to maintain a constant forward velocity. The drag force in this case arises mainly from the viscous shear stresses exerted by the fluid on the body surface; the contributions of "form drag" (resulting from the distal thickening of the boundary layer and possibly from flow separation), of "induced drag" (due to the shedding of vortex sheets), and of "wave resistance" (energy lost through making waves when swimming at the surface) are generally insignificant for these animals.

Thus the hydromechanical problem of fish locomotion can be conveniently divided into two parts: the drag of a streamlined body, analysed by calculating the viscous shear stresses in the boundary layer, and the generation of an equal and opposite thrust, determined from an analysis of the potential flow outside the boundary layer. If the mean thrust is denoted by  $T$ , and the mean drag force by  $D$ , we have, in steady swimming,

$$T = D. \quad (23)$$

#### Power Expenditure and Energy Balance

From the physical standpoint the minimum power necessary to transport the body at a given velocity  $V$  is determined by the resistance of the medium, the efficiency of the mode of locomotion, the energy consumption of the particular type of power plant that delivers the mechanical power, and many other factors. The principle of energy conservation may be expressed as

$$DV = \eta P, \quad (24)$$

which states that the animal's rate of working against the resistance  $D$  is provided by the power input  $P$ , used with efficiency  $\eta$ . For aquatic animals the variations of potential energy are generally unimportant, and in any case can be considered separately as for aerial and terrestrial animals.

The power input for locomotion can also be determined by measuring the animal's metabolic rate and by applying standard energy conversion factors. Thus, when  $P$  in Eqn (24) is taken to be the mechanical power,  $\eta$  is the hydrodynamic efficiency  $\eta_h$  of the propulsion. When  $P$  is taken instead to be the

metabolic rate (the rate at which biochemical energy is released), then  $\eta$  is to  $\eta_h \eta_c$ , where  $\eta_c$  is the efficiency with which biochemical energy is converted to muscle power.

#### Metabolic Rate

The extensive literature on the metabolism and swimming speeds of fish makes it plain that the physical details of an experiment must be clearly specified, in order that its results may be interpreted correctly (see, for example) Winberg,<sup>(15)</sup> Drabkin<sup>(28)</sup> and Brett.<sup>(20, 21, 22)</sup> Of major importance in the study of the metabolic rate of a fish are its level of activity and the temperature of the water (since water holds less dissolved oxygen at elevated temperatures). Such factors as preconditioning (a period of fasting and exercise prior to the test, so that none of the fish's energy is spent in digesting or absorbing food during the test), the state of maturity and sex of the specimen, and flow conditions during the experiment, are also important. We can give only a brief account here.

#### (i) Level of Activity

The level of activity in swimming fish is hard to characterize in general. However, for the salmon, a migratory fish, Brett<sup>(20, 21, 22)</sup> (see Fig. 7) found three levels of performance: sustained (speeds that can be maintained almost indefinitely), prolonged (speeds maintained for 1 or 2 hours with a steady effort, but leading to fatigue) and burst (speeds achieved at maximum effort lasting for only about 30 seconds). The physiological basis of each of these activity levels is different, and itself depends on body size and environmental temperature.

The relationship between swimming speed and metabolic rate can also be evaluated in terms of the rate of working by the muscle, as discussed in other lectures at this Symposium. Lighthill<sup>(47)</sup> reasoned that the efficiency with which muscles produce power increases in proportion to the frequency  $\omega$  of contraction. Hence in both carangiform and lunate-tail swimming  $\eta_c$  should be proportional to the swimming velocity  $V$ . Thus, when the mechanical power required for maintaining the speed  $V$  is proportional to  $V^3$ , the fish consumes energy at a metabolic rate

$$P = P_b + \alpha V^2, \quad (25)$$

where  $P_b$  is its basal metabolic rate and  $\alpha$  is a constant. It therefore follows that  $P/V$ , proportional to the specific energy cost  $\mathcal{E}$  (Eqn 18), is given by

$$P/V = P_b/V + \alpha V, \quad (26)$$

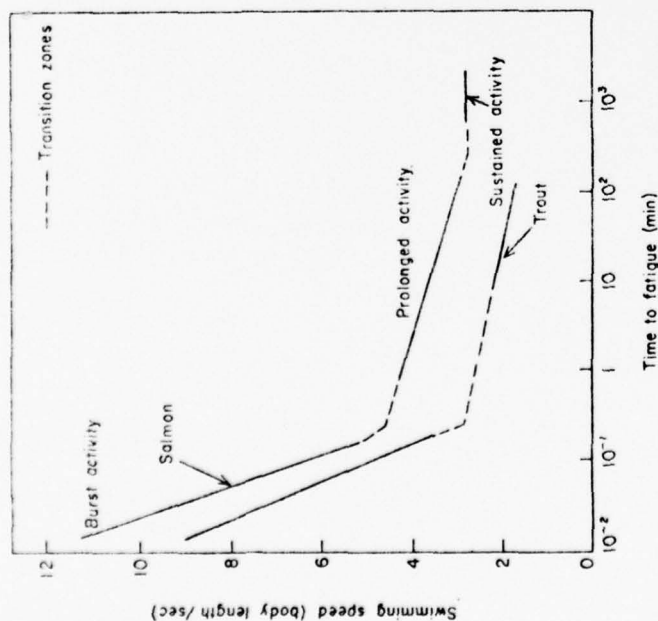


FIG. 7. Swimming endurance of sockeye salmon and rainbow trout. Transition zones between different levels of activity are shown with broken lines. (After Brett<sup>(19,23)</sup>).

which has a minimum at the speed  $V_m = (P_b/x)^{1/2}$ . A speed for which the specific energy cost is a minimum has been found by Brett<sup>(21)</sup> for salmon (see Fig. 8), by Tucker<sup>(64)</sup> for small parrots, and was also noted by Weihs<sup>(74)</sup> in his theoretical study. Figure 8 shows the basic curve of  $\delta$  against  $V$  for sockeye salmon; their ocean migrant speed is very close to the minimum energy cost, and their speeds in Fraser River migration ( $\sim 1.2 \text{ m s}^{-1}$ ) are somewhat lower than their lowest burst speed.

#### (ii) Fish Size and Metabolic Rate

The relationship between body mass and the metabolic rate of fish has received considerable attention, particularly for the resting state (for expository reviews see Kleiber,<sup>(40)</sup> Schmidt-Nielsen<sup>(58)</sup>). The equation which describes this relationship is of the general form

$$P = am^b, \quad (27)$$

(see Eqn 19) or in the logarithmic form

$$\log P = \log a + b \log m, \quad (28)$$

where  $m$  is the body mass, and  $a$  and  $b$  are coefficients independent of body size. Gray<sup>(29)</sup> first tackled this problem by a different approach; he directly considered the total mass of muscle,  $m'$ , so that  $P = a'm'$ , and derived the

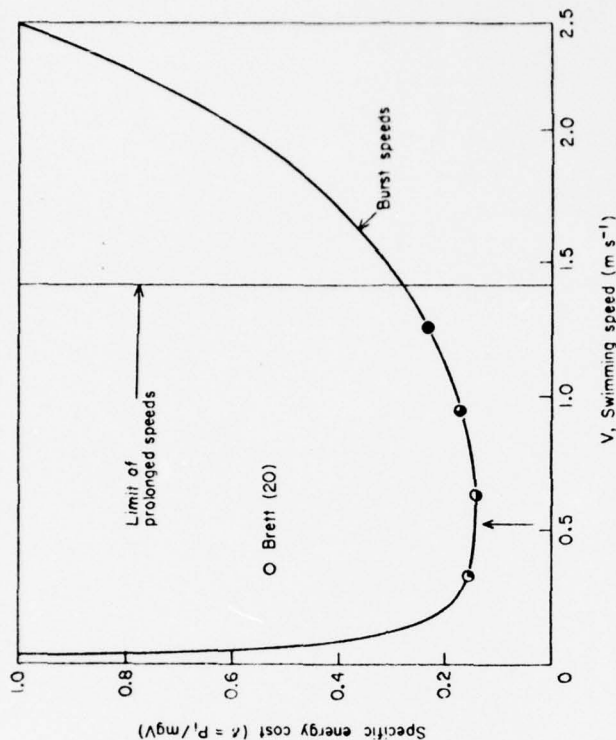


FIG. 8. Specific energy cost  $\delta$  as related to swimming speed, for adult sockeye salmon. Ocean migration occurs slightly above the speed at minimum  $\delta$  (shown with an arrow), which is about  $\frac{1}{2}V_m$  (●), while river migration takes place nearly at  $V_m$  (○). For the definition of  $V_m$  see text and the caption to Fig. 9. (Adapted from Brett<sup>(21,23)</sup>).

power factor  $a'$  relating to unit mass of fish muscle from that of a rowing athlete. Application of this method by Bainbridge<sup>(2,3,4,5)</sup> to several species of fish led to the equation  $m' = 0.005l^{2.9}$ , where  $l$  is the length of the fish in cm when  $m'$  is measured in g.

Various authors have considered body mass  $m$  directly. Winberg<sup>(75)</sup> examined 266 cases for freshwater fish and obtained an average value of  $b = 0.81$  (at  $20^\circ\text{C}$ ). Heusner *et al.*<sup>(34)</sup> investigated three species and derived an overall mean of  $b = 0.73$  (at  $25^\circ\text{C}$ ). There are other reports giving a rather wide range of values, varying all the way from near the "surface law" ( $b = 0.67$ )



to direct proportionality ( $b = 1$ ). The difficulty in seeking a generally applicable value seems to be associated with that met in measuring the drag of a fish.

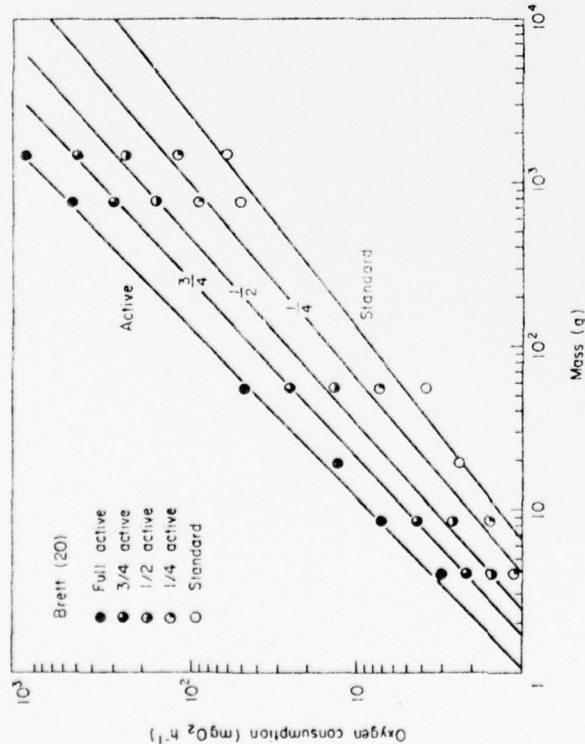


FIG. 9. Metabolic rate of sockeye salmon (*Oncorhynchus nerka*) as a function of body mass at various levels of swimming activity (15°C). The standard level was determined by extrapolation to zero swimming speed of the oxygen consumption vs. speed relation. The line at full activity corresponds to the maximum speed which could be sustained for 60 min,  $V_{crit}$ . Intermediate fraction levels correspond to swimming speeds equal to  $\frac{3}{4}$ ,  $\frac{1}{2}$ , and  $\frac{1}{4}$  of  $V_{crit}$  respectively.

Brett<sup>(18, 20, 21)</sup> has shed considerable and much-needed light on the problem by distinguishing between the different levels of activity. He provided the following results for the sockeye salmon (*Oncorhynchus nerka*), with oxygen consumption measured in  $\text{mg O}_2 \text{ h}^{-1}$ , and body mass in g (see also Fig. 9):

$$\begin{aligned}
 \log a &= -0.632 & b &= 0.775 \pm 0.145 \quad (\text{standard}) \\
 &= -0.523 & &= 0.846 \pm 0.145 \quad (\frac{1}{4} \text{ max}) \\
 &= -0.357 & &= 0.890 \pm 0.145 \quad (\frac{1}{2} \text{ max}) \\
 &= -0.223 & &= 0.926 \pm 0.145 \quad (\frac{3}{4} \text{ max}) \\
 &= -0.050 & &= 0.970 \pm 0.053 \quad (\text{max})
 \end{aligned} \quad (29)$$

Here the fractions are referred to the maximum speed which could be sustained for 60 min in fresh water at 15°C ( $V_{crit}$ ), and his standard metabolic rate is obtained by extrapolating the  $\text{O}_2$ -consumption versus velocity curve to zero swimming velocity. This approach appears to make the study on the scaling of swimming velocity more systematic, as we now proceed to discuss.

### Scaling of Swimming Velocity

In order to use the principles of dynamical similarity, we first express the drag  $D$  on a fish swimming at speed  $V$  in terms of its drag coefficient  $C_D$  as follows:

$$D = \frac{1}{2} \rho V^2 S C_D \quad (30)$$

where  $\rho$  is the water density,  $S$  the fish surface area (proportional to  $l^2$ ) and  $C_D$  depends on the Reynolds number  $Re$  and the body shape. Substitution of this expression and the metabolic rate relationship (27) into the energy equation (24) then yields

$$V^3 \propto l^{3b-2}/C_D \quad (31)$$

Two cases can be examined by assuming that for carangiform and lunatail swimming, in both of which the fish is well-streamlined,  $C_D$  is the same, apart from a shape factor, as that of a flat plate of the same area at the same Reynolds number. That is,  $C_D$  is proportional to  $Re^{-1/2}$  or  $Re^{-1/5}$  according as the boundary layer is laminar or turbulent. This leads to the scaling law:

$$V = \text{const. } l^b \quad (32)$$

with

$$b = \frac{3}{5}(2b - 1) \quad (\text{laminar})$$

$$b = \frac{3}{14}(5b - 3) \quad (\text{turbulent})$$

$$b = b - \frac{2}{3} \quad (C_D = \text{const.})$$

The last case (of quite large but constant  $C_D$ ) corresponds to the situation

TABLE I

| $C_D$                 | $b$ | 0.78<br>(standard) | 0.85<br>( $\frac{1}{4}$ -max) | 0.89<br>( $\frac{1}{2}$ -max) | 0.93<br>( $\frac{3}{4}$ -max) | 0.97<br>(max) | 1.00 |
|-----------------------|-----|--------------------|-------------------------------|-------------------------------|-------------------------------|---------------|------|
| laminar               |     | 0.33               | 0.42                          | 0.47                          | 0.51                          | 0.56          | 0.60 |
| turbulent             |     | 0.19               | 0.26                          | 0.31                          | 0.35                          | 0.40          | 0.43 |
| $C_D = \text{const.}$ |     | 0.11               | 0.18                          | 0.22                          | 0.26                          | 0.30          | 0.33 |



for separated flow past a blunt body with a broad wake formation, or to the case when flow separation occurs in the cross-flow past an undulating fish body. If Brett's values for  $b$  are adopted we obtain Table I for  $\beta$ . The last column, for  $b = 1$ , is included to show the values of the index  $\beta$  when the metabolic rate is proportional to body weight, and the last row, for  $C_D = \text{const.}$ , is for the hypothetical case in which the flow is fully separated.

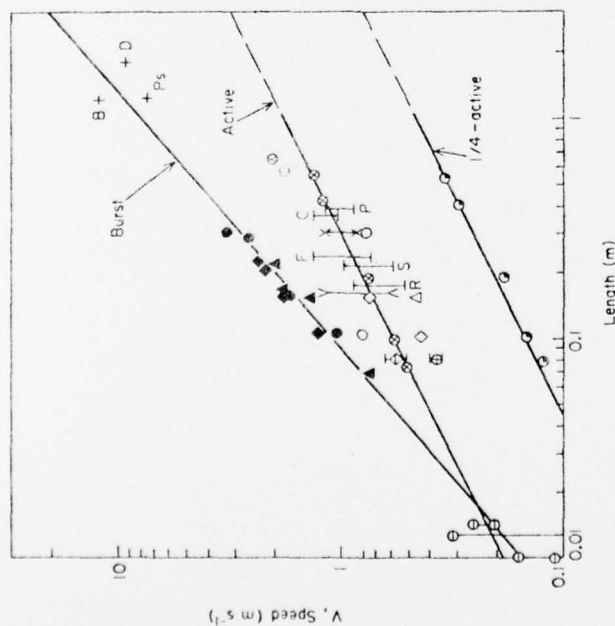


FIG. 10. Variation of swimming speed with body length at different levels of activity. *Burst speed data*:  $\blacklozenge$  dace,  $\blacktriangle$  goldfish,  $\bullet$  trout,  $\circ$  barracuda,  $\square$  porpoise,  $\circ$  dolphin (Bainbridge<sup>(5)</sup>). *Full activity data*: open symbols (Bainbridge<sup>(5)</sup>);  $\otimes$  sockeye salmon (15°C) (Brett<sup>(20)</sup>);  $\odot$  bass,  $\oplus$  coho salmon (Dahlberg *et al.*<sup>(21)</sup>);  $\times$  C cod,  $\Delta$  redfish,  $\nabla$  winter flounder,  $\square$  sculpin,  $\circ$  pout (Beamish<sup>(9)</sup>);  $\triangleright$  goldfish (Smit *et al.*<sup>(50)</sup>);  $\ominus$  larval anchovy (Hunter<sup>(36)</sup>);  $\times$  herring (Jones<sup>(38)</sup>);  $\odot$  salmon,  $\otimes$  trout (Paulick and DeLacy<sup>(55)</sup>);  $\frac{1}{4}$ -activity data:  $\circ$  sockeye salmon at 15°C (Brett<sup>(20)</sup>).

A collection of existing experimental data is given in Fig. 10. In the log  $V$  vs log  $l$  plot for (i) Brett's 60-min. maximum activity results for salmon at 15°C, (ii) Beamish's<sup>(9)</sup> results for 6 species of fish over a range of temperatures (8–14°C) and endurance spans, and (iii) a few other sources (see Fig. 10 caption), the slope of the mean regression line is 0.5 (i.e.  $\beta = 0.5$ ). This result led Brett<sup>(20, 21)</sup> to propose "that the swimming speed is proportional to  $l^{0.5}$ ,

indicating a decrease in the relative ability to maintain a sustained speed as size increases". It is of interest to point out that the speed at the  $\frac{1}{4}$ -max activity level ( $b = 0.85$ ) of the sockeye salmon is also proportional to  $l^{0.5}$  (see Fig. 10). There are, however, other reports (Brett,<sup>(22)</sup> Magnuson,<sup>(50)</sup> Hunter,<sup>(35)</sup> Hunter and Zweifel<sup>(37)</sup>) indicating that a better overall average value for  $\beta$  is 0.6.

Considerable attention has been given to the burst speeds which, for various species of streamlined fish, tend to be nearly independent of both size and temperature. A representative plot is included in Fig. 10, showing Bainbridge's<sup>(5)</sup> results for 4 species of fish and two cetaceans; the slope of this line gives a value of  $\beta$  equal to 0.88. Similar values have been obtained by others, such as  $\beta = 0.94$  for herring (at  $\sim 12^\circ\text{C}$ ) by Blaxter and Dickson,<sup>(16)</sup> In fact, a speed of 10 lengths per second has been regarded by several authors as a "common rule" for the maximum burst speed of many streamlined fish.

At this point a comparison between experiments and the foregoing similitude calculation may have far-reaching implications. First, on the basis of the observed sustained speeds (with the index  $\beta = 0.5$ ) and the corresponding metabolic-rate index  $b = 0.97$ , it may be argued by implication (see the fourth column of Table I) that the boundary layer adjacent to the swimming fish would be at most only partially turbulent up to Reynolds number of  $10^6$  (the upper range covered by the data). Furthermore, there is no evidence to suggest any flow separation. If this argument can be established, the study of the metabolism of swimming fish will yield most valuable results for hydrodynamicists, and may enable them to overcome the difficulties in making measurements in the boundary layer over an undulating body and hence estimating the drag force. Further, we note that the rule of burst-speed  $\approx 10/\text{s}^{-1}$  would correspond to a value of  $b$  equal to 1.33 (for laminar boundary layers), and even higher values when the flow is turbulent. Whether this implies a substantial depletion of stored energy or whether it has some other significance remains to be explored.

### Specific Energy Cost

The results obtained above for the scaling of metabolic rate and swimming velocity may now be used to evaluate the scaling of the specific energy cost,  $\delta = P/mgV$ . From (18), (27) and (32) we immediately deduce that

$$\delta = P/mgV = \text{const. } m^{-\gamma} \quad (33)$$

where

$$\gamma = 1 - b + \beta/3. \quad (34)$$

Using Table I we obtain for  $\gamma$  the following result (Table II):

TABLE II.

| $\beta$ | $\gamma$ | $b$                   | 0.78<br>(standard) | 0.85<br>( $\frac{1}{4}$ -max) | 0.89<br>( $\frac{1}{2}$ -max) | 0.93<br>( $\frac{3}{4}$ -max) | 0.97<br>(max) | 1.00 |
|---------|----------|-----------------------|--------------------|-------------------------------|-------------------------------|-------------------------------|---------------|------|
|         |          |                       | 0.34               | 0.30                          | 0.27                          | 0.24                          | 0.22          | 0.20 |
|         |          | laminar               | 0.29               | 0.24                          | 0.21                          | 0.19                          | 0.16          | 0.14 |
|         |          | turbulent             | 0.26               | 0.21                          | 0.18                          | 0.16                          | 0.13          | 0.11 |
|         |          | $C_D = \text{const.}$ |                    |                               |                               |                               |               |      |

Figure 11 presents the experimental data, derived from those given in Figs 9 and 10, showing how the specific energy cost varies with body size for fish swimming at different levels of activity. The solid lines are regression lines obtained by a least-square-error fit, giving  $\gamma = 0.34$  at  $\frac{1}{4}$ -max, 0.29 at  $\frac{1}{2}$ -max, 0.25 at  $\frac{3}{4}$ -max, and 0.21 at full activity. These values of  $\gamma$  and the observed mean of  $\beta = 0.5$  are plotted against  $b$  (which may be regarded as a measure of the level of activity) in Fig. 12. Also shown are the similarity predictions of  $\beta$  and  $\gamma$  given by (32) (or Table I) and (34) (or Table II) for the three distinct reference states characterized by a laminar boundary layer, a turbulent boundary layer and separated cross flow. This comparison

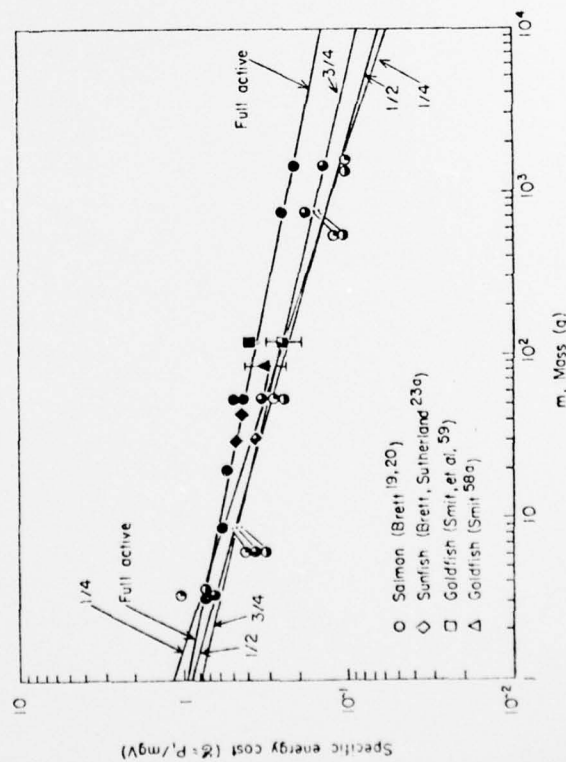


Fig. 11. Relation between specific energy cost  $\bar{C}_D$  and body size for fish swimming at different levels of activity. Solid lines are regression lines obtained from data of Brett<sup>(20)</sup> for salmon at 15°C. For other legend and symbols see Fig. 9.

strongly suggests that the boundary layers of these swimming fish would be largely laminar at these high Reynolds numbers ( $8 \times 10^3 < Re < 1.4 \times 10^6$  being the range covered by the test cases), with at most only a small (presumably distal) part turbulent, and that the specimens tested were swimming at somewhere about  $\frac{3}{4}$  of their maximum sustainable activity.

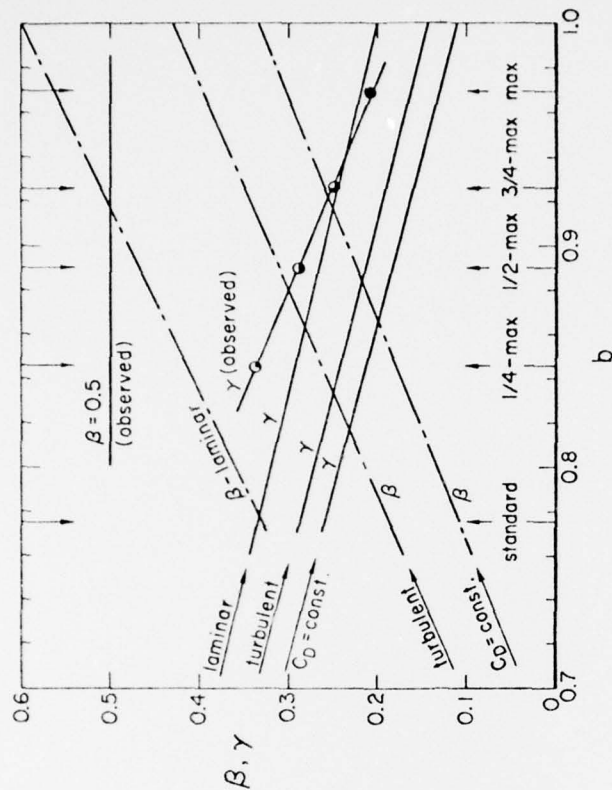


Fig. 12. Comparison of the observed values of  $\beta$  (Brett<sup>(20, 21)</sup>) and  $\gamma$  (see Fig. 11) with the similarity predictions (Eqs (32), (34)) based on the three distinct reference states characterized by a laminar boundary layer, a turbulent boundary layer and  $C_D = \text{const.}$  (for the case of separated cross flows)

The specific energy cost of transport has also been discussed in the recent studies of Schmidt-Nielsen<sup>(57)</sup> and Tucker<sup>(65)</sup> for the locomotion of various animals. For comparison we find, from Schmidt-Nielsen's Fig. 1(a) and Tucker's Fig. 2, that the observed values of  $\gamma$  for fish are

$$\gamma = 0.30 \quad (\text{Schmidt-Nielsen}), \quad (35)$$

$$\gamma = 0.27 \quad (\text{Tucker}). \quad (36)$$

These values of  $\gamma$  are in accord with the present finding when the level of activity lies between about  $\frac{1}{2}$  and  $\frac{3}{4}$  of the maximum performance.

With particular reference to fish size we note from Fig. 11 that the regression lines of  $\delta$  at different levels of activity cross each other at a certain mass. This cross-over suggests that some (specific) energy would be saved if smaller fish were to swim at speeds corresponding to greater activity and larger fish were to cruise at lower relative speeds.

#### Scaling of Tail Beat Frequency and Body Wavelength

Bainbridge<sup>(2,4,7)</sup> observed that the maximum amplitude of the tail beat of fishes in carangiform motion is approximately 0.2 of the fish length  $l$ , with no noticeable dependence on other parameters. The "reduced frequency"  $\omega l/V$ , where  $\omega$  is the radian frequency, is scale independent, and takes values around 10. Observations by other authors are essentially in agreement with Bainbridge's. According to Hunter and Zweifel<sup>(37)</sup> however, the reduced frequency of jack mackerel was found to decrease slightly with increasing size (with a mean value of  $\omega l/V$  equal to 11 for  $l = 4.5$  cm, decreasing to 7.44 for  $l = 27$  cm), while the amplitude remains constant about the value of 0.23  $l$ .

A theoretical estimate of the scaling of  $\omega$  can be obtained by applying slender-body theory (Lighthill<sup>(42)</sup> Wu<sup>(56)</sup> Newman and Wu<sup>(53)</sup> and Newman<sup>(52)</sup>) which provides the following approximate relation for the mean thrust

$$T \propto \rho b_0^2 \omega^2 a_0^2 (1 - V/c), \quad (37)$$

where  $a_0$  represents the amplitude of oscillation of the fish tail,  $b_0$  its characteristic depth, and  $c = \omega/k = f\lambda$  denotes the wave velocity of the wave (of frequency  $f$  and wavelength  $\lambda$ ) which is propagated distally along the body. The dimensionless coefficient multiplying (37) is either constant or very nearly scale-independent. If we assume that  $c/V$  (or, equivalently, the hydro-mechanical efficiency) is scale-independent, equating  $T$  and the drag  $D$  (see Eqn 23) yields

$$\omega l/V = \text{const. } C_D^{1/2} \quad (38)$$

in which we have taken  $a_0$  and  $b_0$  to be proportional to  $l$  and  $S$  to  $l^2$ . Thus, for laminar flow, with  $V$  proportional to  $l^{1/2}$  (see Eqn 32 and its sequel), we obtain

$$\omega l/V = \text{const. } l^{-3/8}. \quad (39)$$

Though this estimate is somewhat crude, it is in qualitative accord with the observed data of Hunter and Zweifel. (The scaling of  $\omega l/V$  would be weaker than  $l^{-3/8}$  for smaller  $\beta < 0.5$ ). The corresponding scaling of  $\omega$  is

$$\omega = \text{const. } l^{-7/8} = \text{const. } l^{-0.88}. \quad (40)$$

Further, if  $c/V$  is again assumed to be scale-independent (which implies that the hydromechanical efficiency is likewise scale-independent), it then follows that the wavelength  $\lambda$  will scale as

$$\lambda = c/f = (V/f)(c/V) = \text{const. } l^{1/8}. \quad (41)$$

It should be of interest to examine these scaling relations when the necessary data are available.

In passing we note that the wing beat frequency of many species of birds tends to be proportional to  $m^{-0.6}$  or to  $l^{-0.78}$  (Greenwalt,<sup>(32)</sup> Alexander (Ref. 1, p. 7)) which is quite close to the expression (40) for fish.

#### Viscous Resistance

As we noted earlier, reasonable accuracy in the measurement of metabolic rate, coupled with direct observations of pertinent parameters such as the fish mass, length, swimming velocity, and the frequency and wavelength of its tail beat, has enabled us not only to carry out the similarity study but also yields valuable information about the drag coefficient. This information appears to contain indirect evidence concerning both the functional dependence of  $C_D$  on the Reynolds number  $Re$  and, even, its order of magnitude, which in turn may throw light on whether there is transition from laminar to turbulent flow in the boundary layer and on the possible occurrence of flow separation in unsteady flows. To acquire such information by direct measurement for freely swimming fish has proved too difficult a task, and it is worth commenting on this.

The great difficulties involved in measuring the drag on a swimming fish directly has forced researchers to assume that it is the same as the drag on an equivalent straight rigid body. Therefore many attempts have been made to measure the viscous drag of either a mechanical model, or a paralyzed or pickled fish, in steady flow in wind or water-tunnels, towing tanks, or water tanks for drop-down tests; another approach has been to observe the retardation of a fish in glide (for brief reviews see Gray<sup>(30a)</sup>, Newman and Wu<sup>(53)</sup>). Generally speaking, the measured values of the drag coefficient  $C_D$  show wide scatter over a range of a few times to tens of times the drag coefficient for turbulent skin friction on a flat plate of the same area at the same Reynolds number. However, there are reports (Lang and Daybell<sup>(41)</sup>, Webb<sup>(73)</sup>) indicating that the measured drag is close to that of an equivalent mechanical model. Drastic improvement in experimental accuracy is urgently needed, but difficult to achieve.

I would like to remark that the basic assumption on the equivalence of swimming drag to model drag may not be generally valid. This is because the pressure field around a swimming fish must differ considerably from that



about a rigid model, and this difference will fundamentally alter the boundary layer calculation. At an appropriate frequency and phase, the undulatory motion may delay both the transition to turbulence and the flow separation.

### CONCLUSION

In the field of aquatic animal locomotion the number of groups of animals which are fundamentally distinct, from either the hydromechanical or the physiological point of view, has been placed by Lighthill<sup>(43)</sup> to be about 10. The Reynolds numbers achieved in their locomotion span the range from  $10^{-6}$  for eukaryotic flagella to about  $10^8$  for the giant blue whale. The different modes of propulsion include the flagellar and ciliary wave motions of protozoa, the undulatory movements in anguilliform, carangiform and lunate-tail swimming of many fish, the flying mode adopted by rays, turtles and penguins, the jet propulsion of squids, and so forth. Equally vast in variety are their distinct physiological characteristics and behavior. Nevertheless, by studying the different scaling problems some quite definite relations between the key parameters have emerged, and these seem to exhibit certain underlying principles. The field is extremely complex, and I have concentrated on only a few points where the problems are relatively easy, and have pointed out some hopeful possibilities for the future. I am quite sure that the conspicuous gaps which I have left in my discussion will be closed by other speakers with their enlightening observations. May I reiterate my conviction that collaborative studies in this multidisciplinary subject can be surprisingly rewarding.

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